

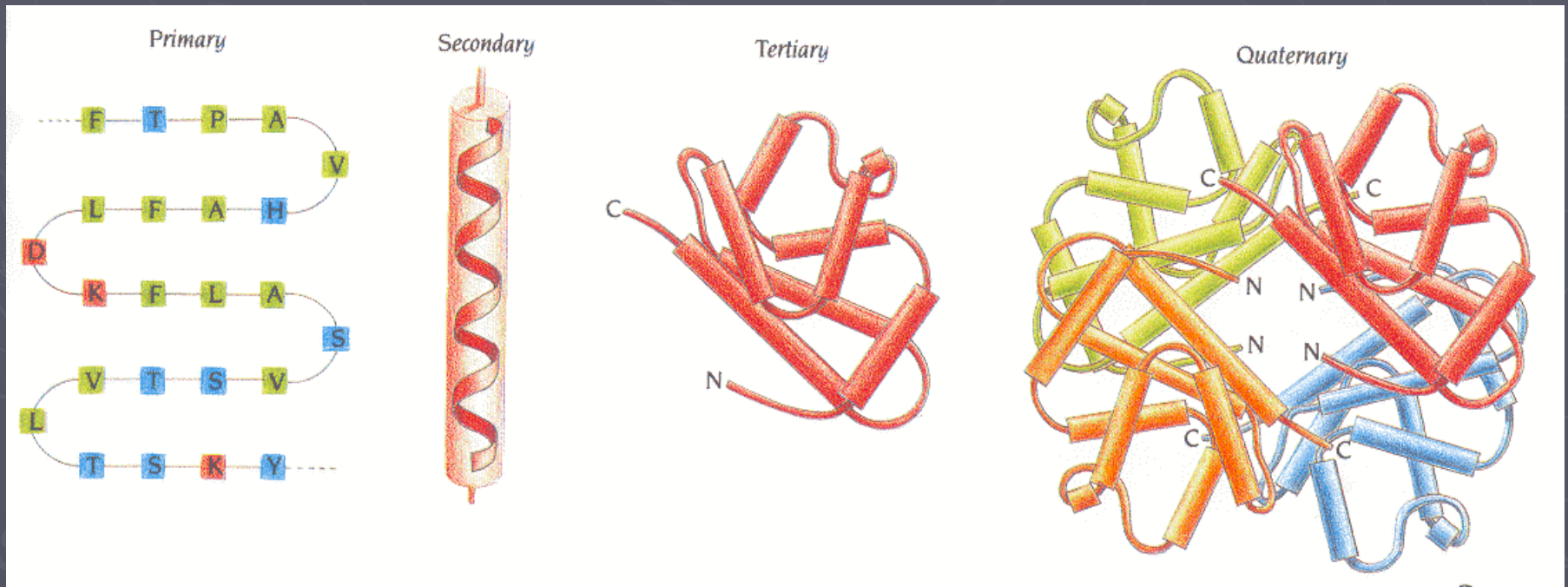
# Structural studies of the methyltransferase NlaX

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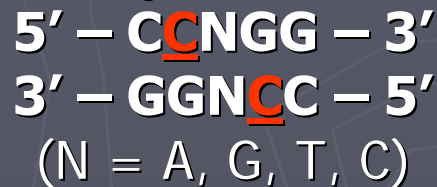
# Protein structures



# NlaX

*Neisseria lactamica*: 34.8 kDa,  
313 amino acid residues  
Expression in *E. coli*: 36.5 kDa,  
6xHis tag on the N-end

C5-cytosine-specific methyltransferase:  
methylates the C5-atom of cytosine in the recognition sequence

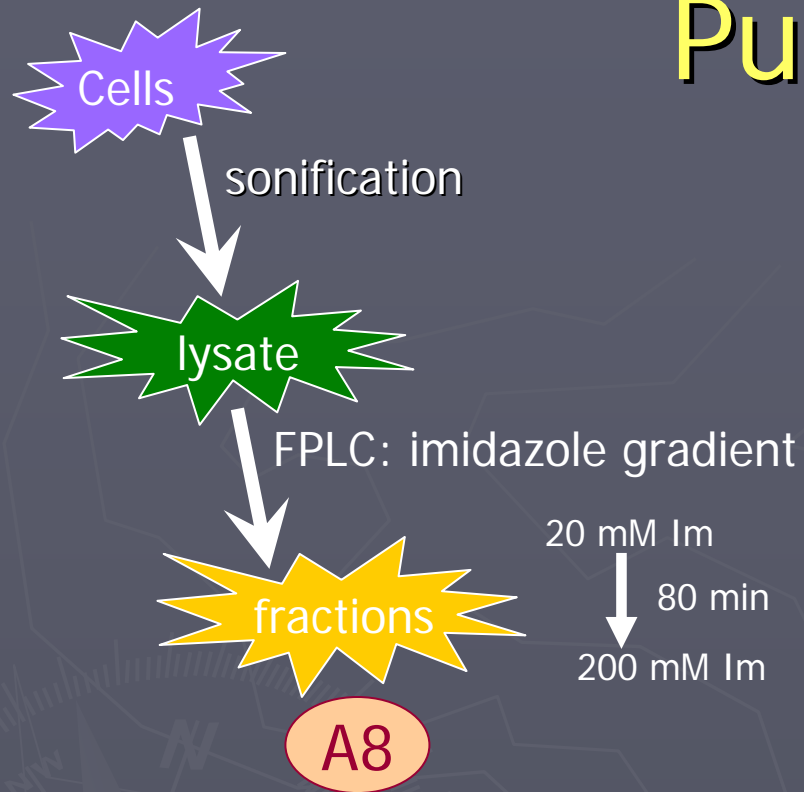


S-adenosyl-L-methionine + DNA

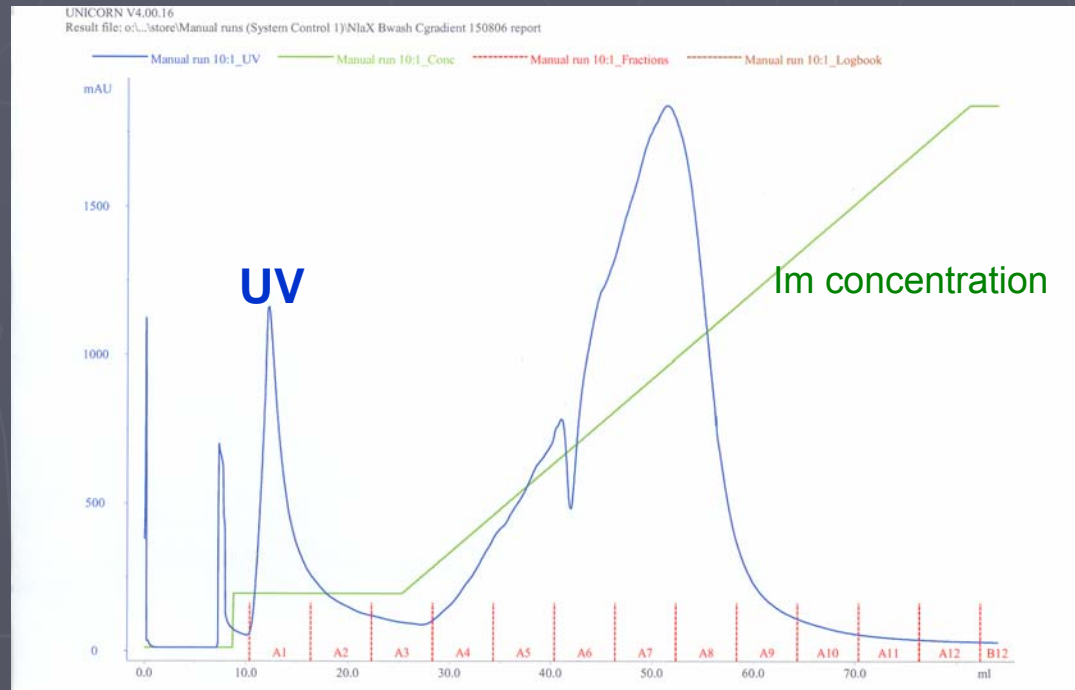


S-adenosyl-L-homocysteine +  
DNA containing 5-methylcytosine

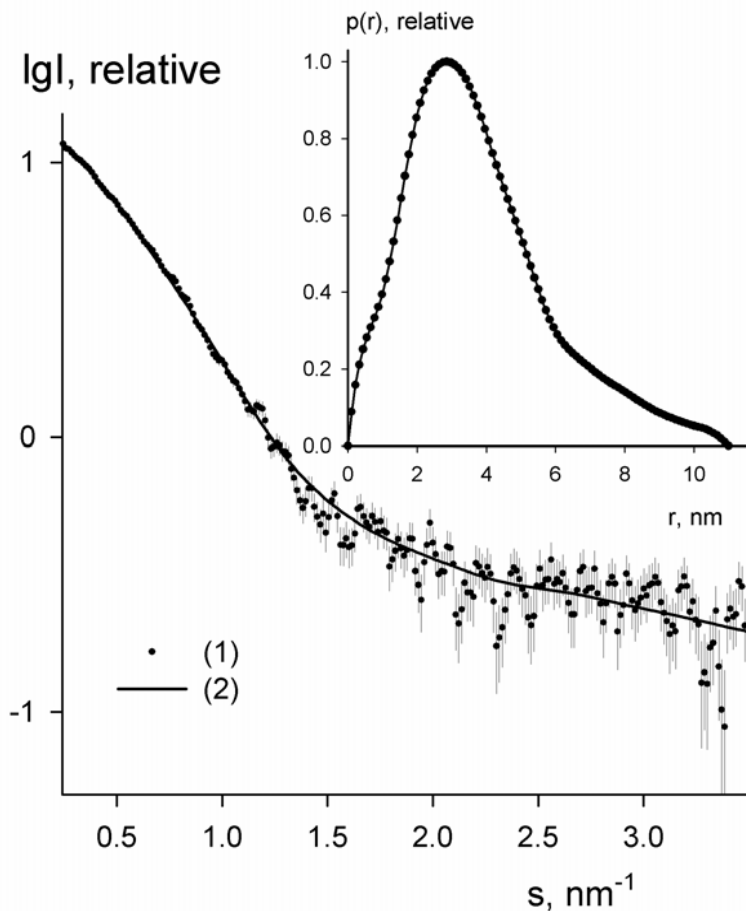
# Purification



A8



A8



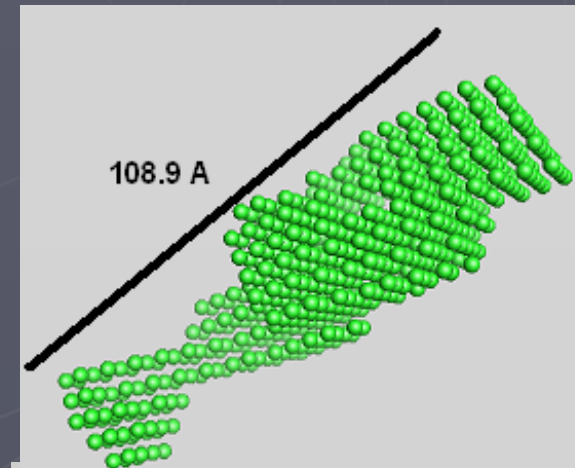
# SAXS

Fraction A8, 10% glycerol

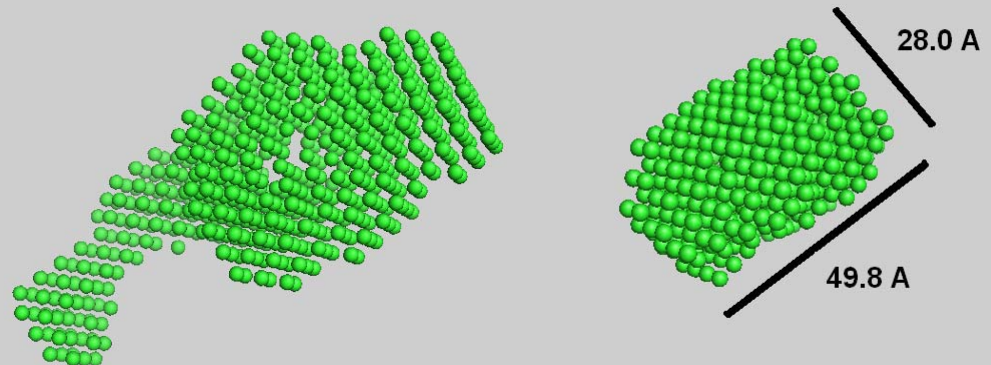
5 mg/ml – spectrophotometer

1 mg/ml – SAXS

**Model**



Radius of gyration	30.42
Maximum diameter	108.9
DAM shape anisometry	0.6274
DAM non-prolateness	0.0
DAM non-oblateness	0.4738



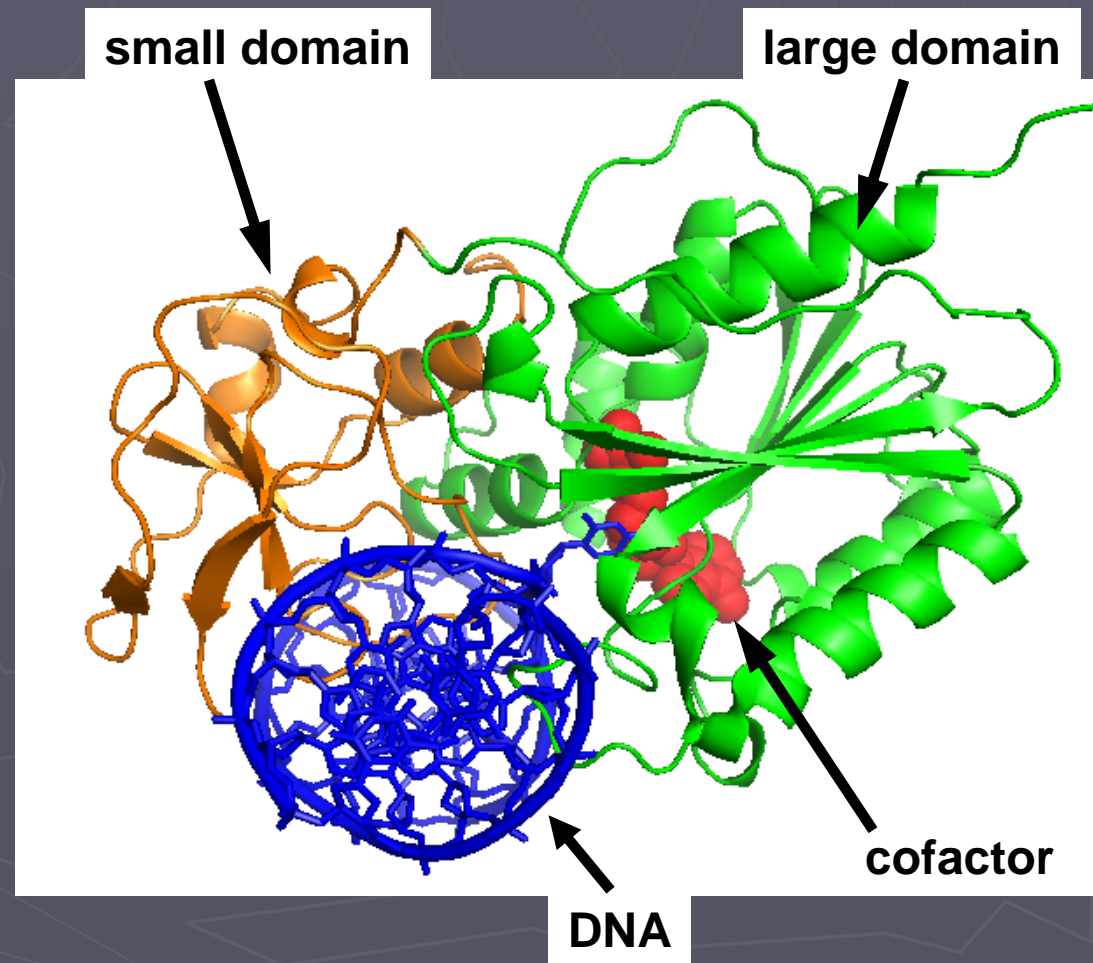
# HhaI – well studied methyltransferase

- ▶ *Haemophilus parahaemolyticus*
- ▶ 327 amino acid residues
- ▶ 37.0 kDa
- ▶ methylates the C5-atom of cytosine in the recognition sequence

5' – GCGC – 3'

3' – CGCG – 5'

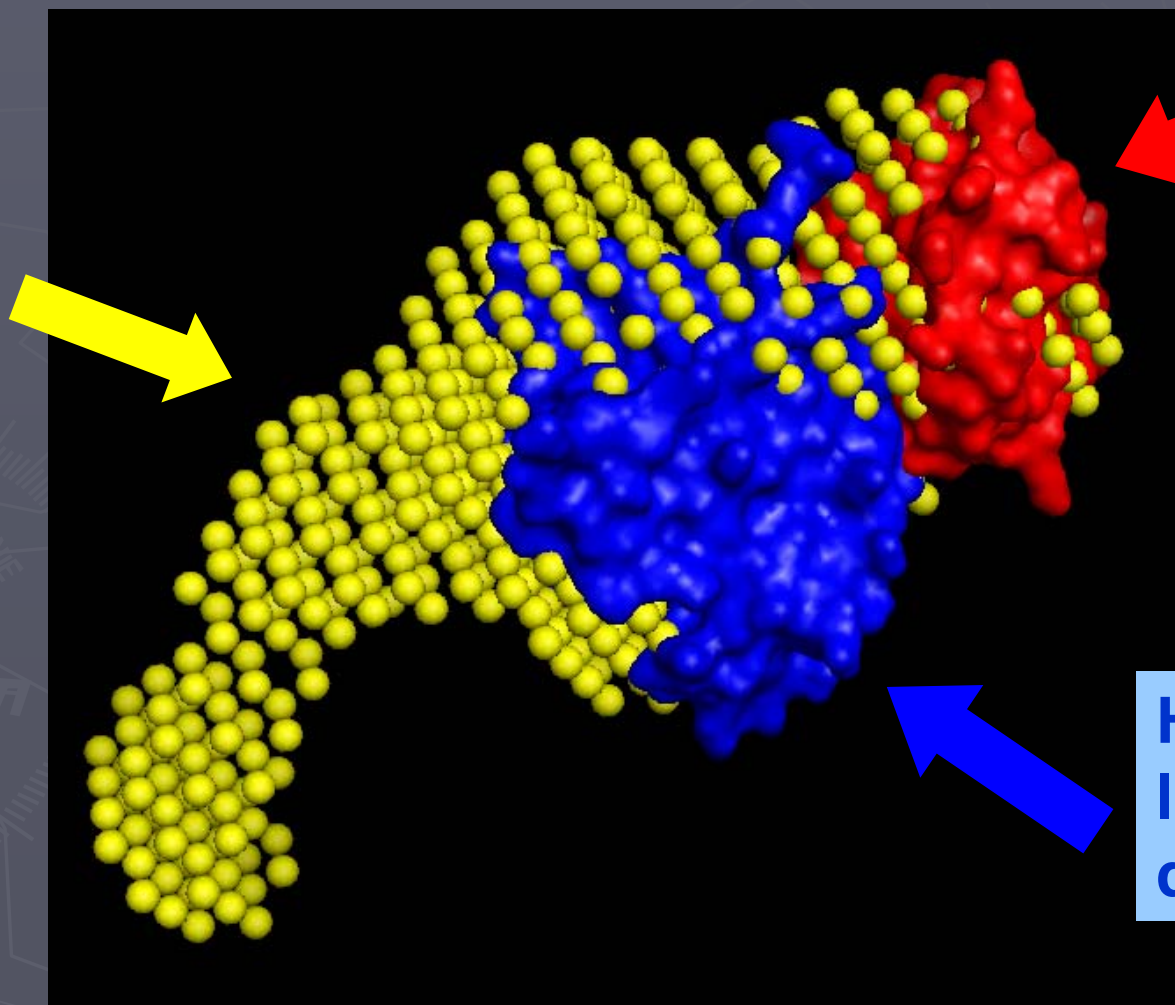
- ▶ Monomer





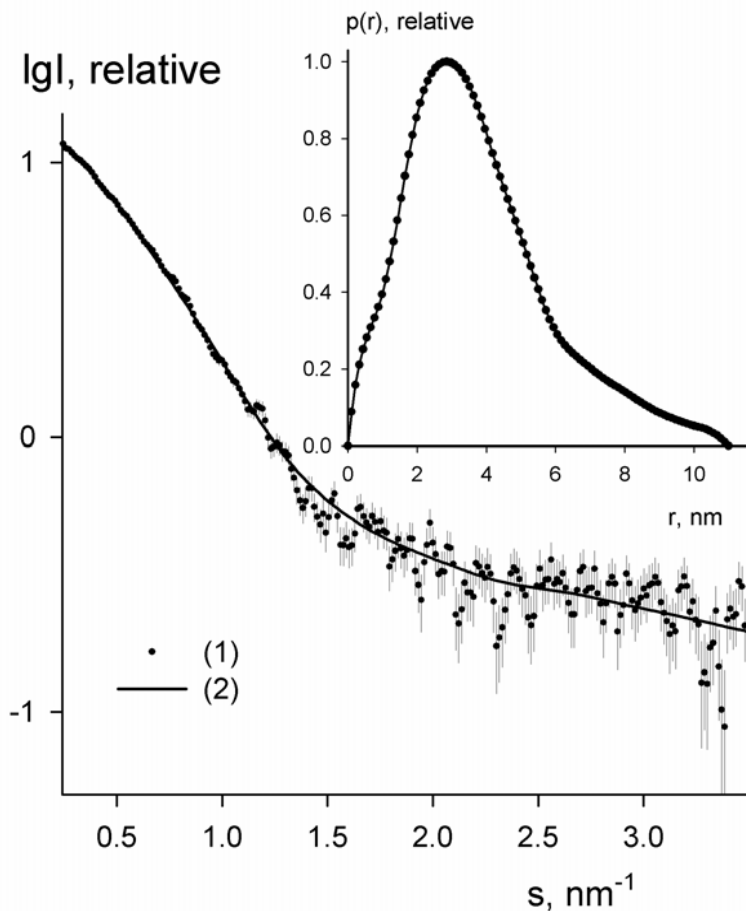
# Superposition

**NlaX  
model**



**HhaI  
small  
domain**

**HhaI  
large  
domain**



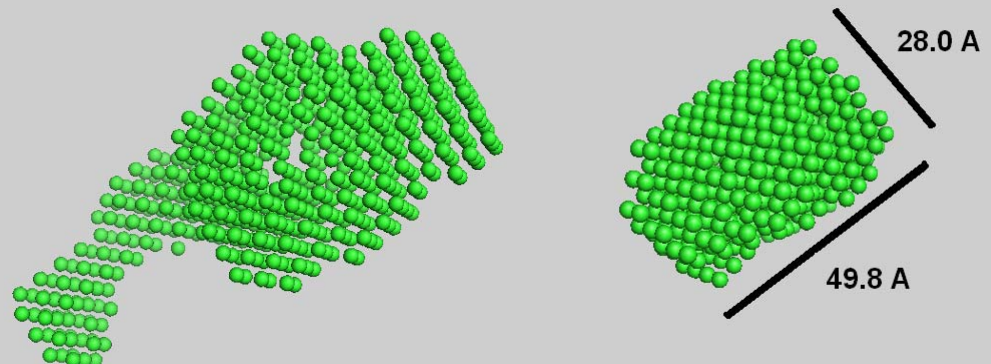
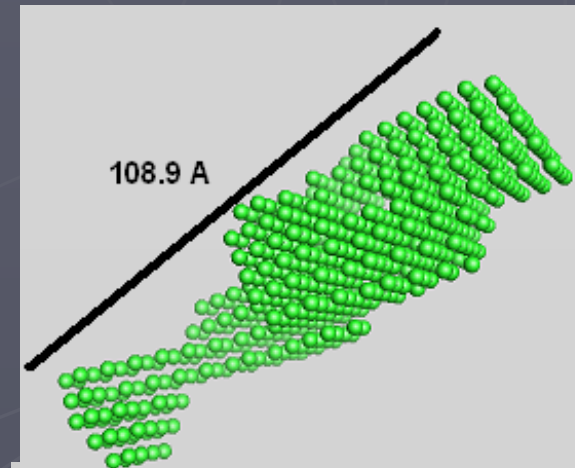
# SAXS

Fraction A8, 10% glycerol

5 mg/ml – spectrophotometer

1 mg/ml – SAXS

**Model**



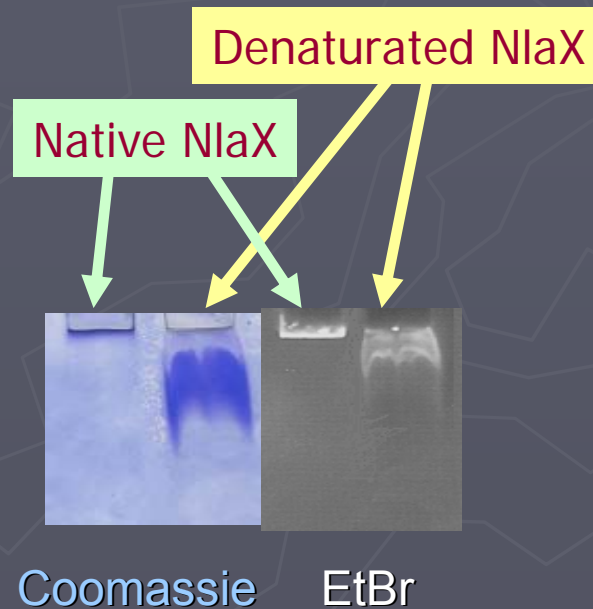
Radius of gyration	30.42
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# NlaX on native gels

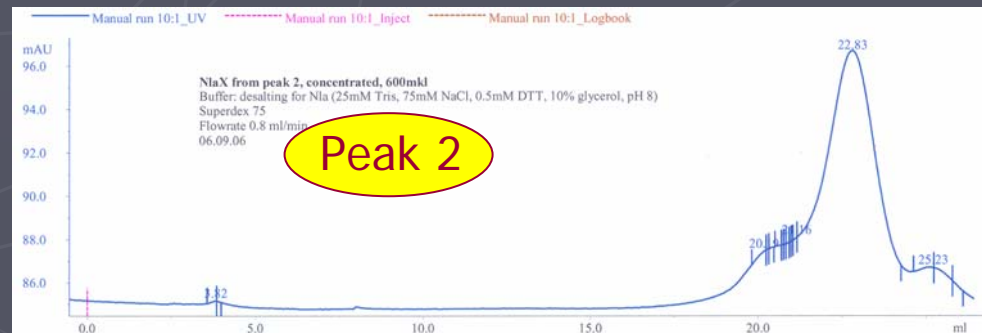
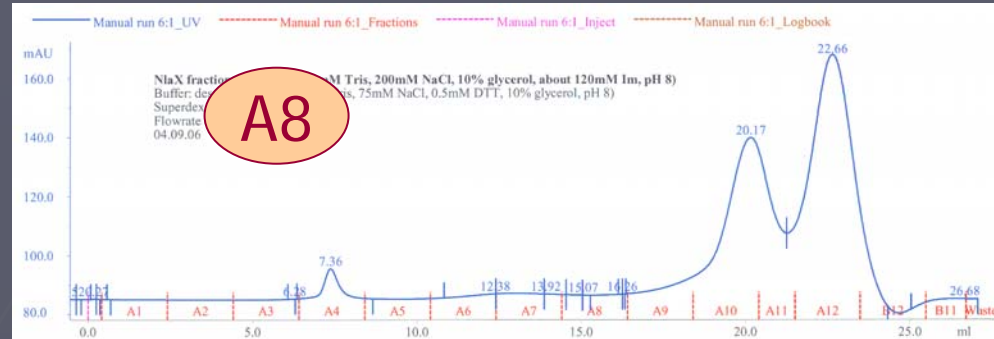
- Native and denatured NlaX:  
staining with EtBr and coomassie

DNA?



# Superdex 75

- ▶ Both peaks contain  $\text{NII}\lambda$
- ▶ No dynamic equilibration between peaks



# Acknowledgements

I would like to thank **Dr. Hans Bartunik** for providing the possibility of this work.

I am especially grateful to **Galina Kachalova**, who proposed important first steps to start the project, explained every aspect of my experiments and helped me with the problems I encountered.

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